

Ocular Biomarkers for Early Detection of Alzheimer's Disease

Shaun Frost^{a,b,c}, Ralph N Martins^{b,d,e}, Yogesan Kanagasingham^{a,c}

^a *Centre for Ophthalmology and Visual Sciences, University of Western Australia, 35 Stirling Highway, Crawley, WA 6009, Australia.*

^b *School of Psychiatry and Clinical Neurosciences, University of Western Australia, Crawley, WA 6009, Australia.*

^c *Commonwealth Scientific and Industrial Research Organisation (CSIRO), Australian E-Health Research Centre, 65 Brockway Rd, Floreat, WA 6014, Australia.*

^d *Sir James McCusker Alzheimer's Disease Research Unit, Hollywood Private Hospital, Nedlands, WA 6009, Australia*

^e *School of Exercise, Biomedical and Health Sciences, Edith Cowan University, Joondalup, WA 6027, Australia.*

Correspondence to:

Professor Ralph Martins, Sir James McCusker Alzheimer's Disease Research Unit, 184 Hampden Rd, Nedlands, WA 6009, Australia.

Tel: (61 8) 9346 6703 Fax: (61 8) 6304 5851

E-mail: r.martins@ecu.edu.au

Running Title: The eye and Alzheimer's disease.

Abstract

Alzheimer's disease (AD) is the most common form of dementia and is characterized clinically by a progressive decline in memory, learning and executive function and neuropathologically by the presence of cerebral amyloid deposits. Despite a century of research, there is still no cure or conclusive pre-mortem diagnosis for the disease. A number of symptom-modifying drugs for AD have been developed, but their efficacy is minimal and short-lived. AD cognitive symptoms arise only after significant, irreversible neural deterioration has occurred, hence there is an urgent need to detect AD early, before the onset of cognitive symptoms. An accurate, early diagnostic test for AD would enable current and future treatments to be more effective, as well as contribute to the development of new treatments.

While most AD related pathology occurs in the brain, the disease has also been reported to affect the eye, which is more accessible for imaging than the brain. AD-related proteins exist in the normal human eye and may produce ocular pathology in AD. There is some homology between the retinal and cerebral vasculatures and the retina also contains nerve cells and fibers that form a sensory extension of the brain. The eye is the only place in the body where vasculature or neural tissue is available for non-invasive optical imaging. This article presents a review of current literature on ocular morphology in AD and discusses the potential for an ocular based screening test for AD.

Keywords:

Retinal, vision disorders, cataract, lens, amyloid beta protein, aging, diagnosis

Contents

Alzheimer's Disease	4
Treatments.....	6
Diagnosis	8
Vision in Alzheimer's Disease.....	11
Ocular Biomarkers for Early Detection of Alzheimer's Disease.....	12
Overview	12
Pupil Responses in Alzheimer's Disease	12
The Ocular Lens and Vitreous Humor in Alzheimer's Disease.....	14
The Retina and Optic Disc in Alzheimer's Disease	17
Conclusions	23
Bibliography	24

Alzheimer's Disease

Alzheimer's Disease (AD) is the most common form of dementia, affecting more than 26 million people worldwide [1]. The disease is characterized clinically by a progressive decline in memory, learning and executive function and neuropathologically by the presence of cerebral amyloid deposits. In addition to the debilitating symptoms endured by AD patients, the disease imposes a huge social and economic burden on society. AD is an incurable, degenerative and terminal disease usually diagnosed in people over 65 years of age [2]. It affects 5% of people aged 65 and 20–40% of those aged 85. The late-onset form of AD (LOAD) is the most common form of the disease and in the majority of cases, where there is no evidence of it being inherited, it is termed "sporadic AD". A rare form of AD, termed "early-onset familial AD" (EOFAD), is inherited in an autosomal dominant manner, and can occur in people as young as 30 years of age. Mild Cognitive Impairment (MCI) is considered a prodromal phase of AD, with 40-60% of people meeting criteria for MCI eventually progressing to AD, or about 5-25% per year [3].

AD is histopathologically characterized by a substantial loss of neurons in the brain, atrophy of the brain, as well as the deposition of extracellular amyloid β ($A\beta$) plaques and intracellular neurofibrillary tau tangles (NFT) [4]. The factors that may cause or accelerate the development of AD are not fully understood, and the exact contribution of plaques and tangles in causing symptoms of AD also remain to be established. General consensus in the field supports the "Amyloid Cascade Hypothesis", which states that an imbalance in $A\beta$ metabolism in the brain is the fundamental cause of the neurodegeneration and cognitive decline in AD, though many studies suggest this effect of $A\beta$ is at least partly mediated by increased tau phosphorylation. In recent years, much evidence has been gathered to show that several factors contribute to the risk of developing AD. These include diabetes, mid-life

obesity, and a history of heart disease or symptoms typically associated with heart disease such as high levels of low density lipoproteins (LDL) together with low levels of high density lipoproteins (HDL) [5].

The major protein component of the amyloid plaques is a peptide known as amyloid β ($A\beta$). $A\beta$ peptides range from 39 to 43 amino acid residues in length. The longer ($A\beta_{42}$ or $A\beta_{43}$) peptides aggregate easily into fibrils, and small soluble oligomers of $A\beta$ are believed to be a neurotoxic form of $A\beta$, whereas the large insoluble aggregates and plaques are relatively inert [6]. The $A\beta$ peptide is proteolytically derived from its parent molecule, the amyloid- β protein precursor ($A\beta$ PP). $A\beta$ PP is an integral membrane protein that has been implicated in blood clotting [7] and as a regulator of neural plasticity and post-injury repair [8]. $A\beta$ peptides are produced as a result of sequential cleavage of $A\beta$ PP by enzymes known as the β -secretase (also known as β -site $A\beta$ PP cleavage enzyme or BACE) and γ -secretase, respectively. The most common form is $A\beta_{40}$, but it is the second most common form, $A\beta_{42}$, which is more fibrillogenic and is thus associated with disease states [9].

$A\beta$ has been shown to have a constrictive effect on the cerebral vasculature [10], and interestingly, to be neuroprotective at low physiological concentrations [11]. For this reason $A\beta$ has been suggested to have a dual damage response role in the brain, by sealing the vasculature reducing brain oxygen requirement and combating oxidative stress [12]. A recent study also demonstrated that $A\beta$ has significant antimicrobial ability against clinically relevant organisms, introducing an interesting hypothesis that infection might have a role in some forms of AD [13]. Whatever the role of $A\beta$ in the healthy body, a large number of *in vitro* and *in vivo* AD studies, particularly those of EOFAD, have shown that higher than normal $A\beta$ levels cause oxidative stress in the brain, resulting in synaptic loss, cell membrane damage, inflammation and ultimately result in neuronal cell death [14-15]. Such

studies have shown that EOFAD-associated mutations in the A β PP gene or the presenilin 1 and 2 genes (presenilin being an essential component of the γ -secretase enzyme) have been found either to increase total A β levels or to increase the production of A β 42 [16-17]. Such increases in A β levels have been implicated in the pathogenesis of both familial and sporadic AD [14-15].

NFT are insoluble, twisted fibers of a protein called tau. Phosphorylated tau stabilizes the internal structure of healthy neurons, but in AD, tau proteins are hyperphosphorylated and tangle together. Again, recent evidence points to soluble tau rather than NFT as a cause of neuronal loss in AD [18-20]. When researchers turned off the mutant tau gene in a mouse model of AD, neurodegeneration was halted and cognitive performance improved even though NFT continued to accumulate [19]. While tau hyperphosphorylation is not specific to AD, there is strong evidence to indicate that it is essential for A β -induced cognitive decline to occur.

Treatments

At present there is no cure or disease modifying drug to effectively treat AD, but there are drugs that have proven to be beneficial at the level of reducing some symptoms for up to 18 months. There is an urgent need for effective treatments, with the number of cases worldwide forecast to exceed 100 million by 2050 [1]. Currently available drugs can delay or alleviate symptoms but do not slow the progression of the disease as they do not target the underlying cause. Pharmaceutical drugs currently available to treat the cognitive manifestations of AD include acetylcholinesterase inhibitors and an NMDA receptor antagonist. Research is continuing into drugs designed to prevent or reduce A β production,

to break up A β plaques or to prevent metal ion-A β interactions which may accelerate A β aggregation [21-22]. For example, one suggested avenue of treatment involves modulating the activity of β and γ -secretases to produce mainly A β 40 instead of the more harmful A β 42 peptide. In other studies, immunizing transgenic AD mouse models with human A β 42 peptide was found to prevent the build-up of A β plaques [23] which was associated with prevention of memory impairment, yet a successful outcome was not achieved in early clinical studies due to serious side-effects. However, immunotherapies that either prevent plaque deposition or enhance removal of plaques are still being actively investigated by pharmaceutical companies with a number of them in either phase II or III clinical trials. Anti-aggregation agents with the aim of removing plaques and/or preventing A β fragments from aggregating are also being examined [24] and are expected to enter clinical trials within the next 12 months.

In 2008, a discontinued Russian antihistamine drug *Dimebon (latrepirdine)* was found to stabilize cognitive and functional activity in AD patients [25]. However, this phase II study outcome has since been thrown into question by negative results from subsequent phase III trials [26]. Differences between the studies include mean age and cognitive performance of participants as well as the source of Dimebon. Further studies are now being conducted to determine the efficacy of Dimebon in conjunction with other drugs to treat AD. Dimebon inhibits cholinesterase and NMDA receptors and may enhance neuronal function and survival through its effects on mitochondria. Other non-pharmacological therapeutic approaches are currently being investigated. Of these, lifestyle modification, particularly physical activity, nutrition and mental stimulation are gaining considerable attention in the field. It has been demonstrated that a cognitively stimulating environment can reduce A β deposition in transgenic mice [27], and that increased cognitive activity in humans can

reduce the risk of AD [28]. A possible explanation for the mechanism by which cognitive stimulation might delay onset of AD has been identified [29]: researchers found that synaptic activity increases A β in local interstitial fluid. Cognitive stimulation transfers electrical activity away from brain regions with high 'default activity', hence reducing A β levels in these regions – particular regions which in fact end up with high levels of A β deposition in AD [30].

The above-mentioned treatments can be made available for AD patients only after they have been clinically diagnosed with the disease. Unfortunately AD is difficult to diagnose with absolute certainty. In addition, the cognitive symptoms on which diagnosis is based only become apparent after irreversible brain damage has already occurred. Hence the search for better AD treatments needs to be coupled with research into early detection of the disease.

Diagnosis

There is currently no definitive *pre-mortem* diagnosis for AD. Conclusive diagnosis of AD is only achieved following *post-mortem* examination of the brain for the presence plaques and tangles in the brain. A pre-mortem diagnosis of "probable AD" is currently made by clinical observations and testing of cognitive capacity and memory loss. Other dementias and conditions such as depression can have similar symptoms, thus confounding diagnosis [31]. A diagnostic error rate of about 10-15% has been reported for AD [32].

A diagnosis of probable AD is only possible when the condition has progressed and considerable neurological damage has already occurred. The increasing frequency of AD in the population, along with the need to treat the disease before cognitive symptoms arise,

calls for a sensitive and specific screening technology to identify high risk individuals before the brain is irreversibly damaged.

Biomarkers that have shown promise for the early detection of AD include a reduction in brain and specifically hippocampal volume (using MRI – magnetic resonance imaging), changes to brain function (using functional MRI) and changes in the concentrations of A β 40, A β 42 or tau in the cerebrospinal fluid (CSF). CSF biomarkers have proven to be the most accurate of the biomarkers investigated to date, with AD being shown to be associated with decreased A β 42 levels and increased tau levels in the CSF [33]. However the latter procedure is invasive and patient compliance is anticipated to be low.

Another promising approach is PET imaging employing ligands, such as Pittsburgh Compound-B (PIB), which selectively bind to A β plaques *in vitro* and *in vivo*, enabling plaque load to be imaged in living patients [31-33]. Interestingly, many MCI and a significant percentage (30%) of cognitively normal elderly individuals show high brain retention of PIB and low A β 42 levels in the CSF [34-35]. Further follow-up studies are required to determine if this identifies those individuals that will go on to develop AD, with one recent study supporting this hypothesis [36].

As mentioned earlier, in terms of genetic markers, certain mutations in A β PP, presenilin 1 and presenilin 2 are known to cause EOFAD [16-17]. One major genetic risk factor for sporadic AD has been known for some time, the apolipoprotein E (APOE) ϵ 4 gene [37]. APOE ϵ 4 has been implicated in modulating the metabolism and aggregation of A β [38]. Individuals with one copy of the APOE- ϵ 4 allele have a 2-3 fold increased risk of developing AD by age 85 and those with two copies have a 12 fold increased risk compared to the general population [39].

Recent genetic association studies have revealed three more genes linked to AD, the Clusterin, PICALM and CR1 genes [40-41]. Also, a recent meta-analysis has found more than 20 genes that have a significant effect on AD risk [42]. However, while these genes may contribute to identifying high risk individuals, they are not sufficient on their own to make a diagnosis, nor is the major genetic risk factor, APOE ϵ 4, which is associated with up to 50% of all AD cases. Hence genetic profiling is emerging as a technique for predicting the *risk* of an individual getting AD rather than providing diagnostic markers. It is possible that a panel of genetic and biological factors may together prove useful in determining high risk individuals.

Important properties of a biomarker for screening are sensitivity and specificity for the relevant disorder, as well as practicality of the biomarker measurement. Evaluation of the sensitivity and specificity of AD biomarkers involves comparison of biomarker measurements in AD cases and controls. Evaluation of potential AD biomarkers is thus hampered by the 10-15% diagnostic error rate in AD [32]. Studies incorporating post-mortem confirmation of AD can avoid this problem, but for most studies this is not possible, hence this diagnostic inaccuracy should be considered when interpreting results. A significant number of healthy controls (30%) may be high-risk clinically silent individuals, further confounding interpretation [43-44].

The combination of different biomarkers can increase sensitivity and specificity; combining CSF A β 42 and tau parameters increases sensitivity and specificity above 80%. Indeed Simonsen *et al.* [45] found a panel of 5 CSF biomarkers for AD which when combined gave 100% sensitivity and 97% specificity on blinded independent data [45]. However, this study compared AD cases with healthy controls and hence doesn't provide information about the specificity of the test against other dementias or diseases. In terms of practicality, lumbar

punctures for CSF fluid samples are expensive, invasive and patient non-compliance is a major issue. Neuro-imaging (MRI, PET, CT) is also expensive and the number of facilities available for clinical use is not adequate for population screening. Therefore there is a need for new AD biomarkers that are more practical as well as more sensitive and specific. The absence of a suitable screening technology for AD has motivated some researchers to look for biomarkers that might exist elsewhere in the body, including the eye.

Vision in Alzheimer's Disease

Visual disturbance is often an early complaint of AD patients [46-47] and studies have reported reduced visual performance on tests of visual field [48-49], color vision [50-52], contrast sensitivity [53-55], backward masking [56-57], visual attention, motion perception, shape-from motion, visuo-spatial construction, visual memory [58-60], delayed saccadic initiation and movement and fixation problems [47, 61-63]. However, none of these deficiencies are specific to AD. The current literature is controversial and reflects the need for larger and more rigorous studies to be undertaken before the significance of this interesting approach for AD-screening can be conclusively evaluated.

Reported visual deficits in AD have generally been attributed to neuronal damage in the visual pathways of the brain [64-65] as well as deficiency of the neurotransmitter acetylcholine in AD, which is important in visual processing [66-68]. Indeed there is evidence that during the pathogenesis of AD, plaques and tangles occur in visual processing brain regions prior to their occurrence in the hippocampus [65]. Thus visual disturbance in AD may precede memory impairment. Since these visual deficiencies are not specific to AD, a newer field of research is investigating the hypothesis that there might be specific

pathological changes in the eye that accompany the disease. Such ocular changes may contribute to the visual deficiencies, or be a result of damage to the visual pathways of the brain. If the eye does harbor an endophenotype of AD, this would give hope for an ocular diagnosis of AD as well as opening up a new avenue for finding other genetic determinants of the disease. There is hope that the eye might yield biomarkers that are either highly specific for AD, or can contribute to an AD-specific risk-profile analysis in combination with genetic, cognitive and other tests. The following sections describe reported AD-associated changes to the eye.

Ocular Biomarkers for Early Detection of Alzheimer's Disease

Overview

Statistically significant ocular abnormalities that have been reported to accompany AD are tabulated below and discussed in the following sections;

(Table 1)

(Figure 1)

Pupil Responses in Alzheimer's Disease

The pupil is the aperture stop of the eye and controls the retinal illumination (see Figs. 1 and 2). The size of the pupil is regulated by the brain in response to the signals it receives from the eyes. Pupil size changes with brightness of incident light, emotions (e.g. fear), pain, cognitive tasks and with use of certain drugs (e.g. alcohol, opioids, LSD).

(Figure 2)

Pharmacological drugs delivered in eye-drops can affect the iris muscles, causing the pupil to contract or dilate, as is often required for ophthalmological examinations. A hypersensitive pupil response to a cholinergic agonist (*pilocarpine* - contraction) or antagonist (*tropicamide* - dilation) has been reported in AD patients [69-78]. The neurotransmitter acetylcholine is deficient in the AD brain [67, 79], hence cholinergic dysfunction in iris nerve cells is a possible explanation for the hypersensitive response, although in this case one would expect to find agonist hypersensitivity with antagonist subsensitivity, or vice versa. Alternative explanations more consistent with the dual hypersensitivity include AD related damage to the locus coeruleus brain region which is involved in pupillary control [80] or increased corneal penetration of the cholinergic eye-drops. One study using a fluorescent marker to evaluate corneal penetration of tropicamide found no difference between AD and controls [81], but further studies are required to confirm this result. It should also be noted that not all studies controlled for medications with anticholinergic effects.

The hypersensitive response was in some cases identified early in the disease progress, encouraging utilization for AD screening, however other studies have brought into question both the sensitivity and specificity of the test. Some studies found no significant hypersensitivity in AD [81-91], or a hypersensitive response in APOE- ϵ 4 allele carriers rather than AD [78, 92], affecting carriers of this allele who are cognitively normal (although at increased risk of progressing to AD). A similar hypersensitivity has been reported in Down's syndrome subjects – who develop AD as a result of an extra copy of the A β PP gene [93], yet also in healthy young adults [87]. Still other studies have demonstrated modulation of the

pupil dilation response by eye color [93] or age [83-84, 94]. These results have left the reliability of the pupil dilation test for AD in question.

The pupil light reflex (Fig. 2) is the response of the pupil to a bright flash of light, involving rapid contraction followed by dilation back to original size. This reflex has been used as a neurological screening tool for disorders such as Parkinson's disease, Huntington's disease, schizophrenia, multiple sclerosis and trauma [95-98]. Since it is mainly a parasympathetic cholinergic response [99], the pupil light reflex could also possibly be affected if central cholinergic depletion in AD extends to the parasympathetic oculomotor system.

The pupil light reflex has been investigated as a non-invasive, ocular predictive marker for AD. Changes to a number of response parameters have been found in AD compared to healthy ageing [100], with a single parameter (reduced "maximum constriction acceleration") facilitating perfect classification in one study. This built upon previous studies which also found statistically significant differences in pupil flash response between AD and controls [91, 101-102], but Granholm *et al.* [91] found that AD and Parkinson's disease patients exhibited the same results. This suggests that specificity might also be an issue for this test, although "maximum constriction acceleration" was not considered in this study. The pupil flash response is also influenced by age [103]. More research is required to confirm the sensitivity and specificity of the pupil flash response parameters as a screening tool for AD.

The Ocular Lens and Vitreous Humor in Alzheimer's Disease

When light enters the eye it passes through the outer 'corneal' layer, followed by the aqueous humor and then the intra-ocular lens (see Fig. 1). The role of this anterior region of

the eye is to focus an optical image of the outside world onto the retina. One disorder that can disrupt this role is *cataract*; an opacification of the lens often due to protein aggregation (see Fig. 3). Cataracts are a common problem in the elderly, with progressive deposition of insoluble protein in the lens and extensive oxidative damage generally caused by environmental factors such as UV exposure [104-105].

The ability of the lens to focus light is achieved by its high protein concentration, higher than any other tissue of the human body. This high protein concentration, along with the optical accessibility of the lens, makes it ideal for the optical investigation of protein aggregation in disease, *in vivo*. The A β protein involved in the pathogenesis of AD in the brain has also been found to exist in the lens (A β 40 and A β 42), aqueous humor (A β 40) and vitreous humor (A β 42) of the normal human eye [106-107].

(Figure 3)

Remarkably, research indicates that a particular type of cataract (equatorial supranuclear cataract) might be specific to AD sufferers [106]. Slit-lamp microscopy of *ex-vivo* intra-ocular lenses of individuals with AD consistently revealed equatorial supranuclear cataracts. Subsequent histochemical analysis in the same study indicated that A β aggregates are present in the cytosol of the lens fiber cells co-localizing with the cataracts. This cataract has also been reported in Down's syndrome subjects [108] – who develop AD as a result of an extra copy of the A β PP gene.

These cataracts (or the initial A β aggregation in the lens) could thus be a biomarker for AD, although it is unknown at which stage of AD pathogenesis they occur. The location of the AD cataract is at the equatorial periphery of the lens posterior to the iris. The anatomical relationship of the lens relative to the iris renders AD-linked supranuclear opacification

virtually harmless with respect to visual impairment and difficult to detect on routine physical examination. However, these AD-linked lesions are readily observed by slit lamp ophthalmological evaluation in fully dilated subjects. If A β is indeed aggregating in the AD lens, leading to these cataracts, it is possible that the initial molecular changes could be detected non-invasively as an early screening or diagnostic test. Further research is needed to establish the specificity of these cataracts and lens A β aggregations to AD, since both A β PP and A β have been shown to increase in concentration in the normal mammalian lens in response to UV radiation or other oxidative effects [109].

For early diagnosis of AD, it has been proposed that a suitable eye-drop biomarker may enable A β in the anterior chamber to be stained and quantified [106]. Alternatively, the size of A β aggregates in the eye may facilitate non-invasive detection with optical scattering [110], spectral or autofluorescence techniques. Dynamic light scattering (DLS) uses back-scatter from a low energy laser beam to determine information about particle size, shape, movement and interactions. The technique is applicable to all eye tissues and has already shown promise for early cataract detection by monitoring the α -crystallin proteins that prevent protein aggregation in the lens [111-113].

Raman spectroscopy and AF techniques both involve illumination of the sample at a specific wavelength followed by measurement of the in-elastically scattered light at different wavelengths. Raman spectroscopy with principal components analysis has been used to distinguish AD from control *ex vivo* post-mortem brain tissues, based on spectra of protein aggregates [114-117]. AD brain tissues also exhibit visual and infrared-excited AF [116, 118]. These techniques could prove useful for non-invasive, early diagnosis of AD using the eye.

The eye contains three main fluid chambers called the anterior, posterior and vitreous chambers. The aqueous humor fills the anterior and posterior chambers (see Fig. 1),

providing nutrients to the lens and cornea and maintaining the convex curvature of the cornea. The vitreous humor has functional interactions with the lens and retina. No changes in these chambers have been reported in AD to date, but a change in A β 42 and tau protein levels in the vitreous humor has been linked to retinal diseases such as diabetic retinopathy and “glaucoma concurrent with other ocular diseases” [107]. This change in protein levels is similar to that observed in the CSF in AD. Given the retinal degeneration observed in AD [119-123] and the recently reported common features between AD and glaucoma [61, 107, 121-130], the vitreous humor is an interesting focus for future research into ocular protein changes in AD.

The Retina and Optic Disc in Alzheimer’s Disease

While the role of the anterior eye is to focus light onto the retina, the retina’s task is to convert the light into electrical signals that enter the brain. The retina consists of multiple layers of neural and photoreceptor cells, along with nerve fibers and vasculature (see Figs. 4-6). The optic disc (or optic nerve head) is the interface between the retina and the optic nerve and is the location at which blood vessels and retinal nerve fibers leave the retina. Reduced visual performance in AD may be the result of pathology in the visual centers of the brain, but it also remains a possibility that retinal degeneration is involved, perhaps as a consequence of visual centre damage.

Ocular morphology reported in AD includes changes to the retinal vasculature and optic disc, retinal cell loss and thinning of the retinal nerve fiber layer (RNFL). A recent study by

Berisha *et al.* found that AD participants had a specific pattern of RNFL loss (measured by optical coherence tomography, OCT), narrower venules and decreased blood flow in these venules (both measured by a laser Doppler instrument) [120]. A limitation of the study was the small participant numbers (9 probable AD and 8 controls). It should also be noted that retinal vessel widths are influenced by age and race and can be altered in many disorders [131]. Decreased retinal venular caliber (vessel diameter), as reported in AD, has otherwise only been associated with high current blood pressure and low high-density lipoprotein (HDL) cholesterol levels [131]. In contrast, *increased* retinal venular caliber is associated with hyperglycaemia, obesity and inflammation [131].

(Figure 4)

If retinal vascular constriction is associated with AD, it is unclear whether the reduced blood flow might be responsible for the reported retinal cell death or instead might be a response to the associated reduction in metabolic demand. A β has been reported to exhibit a constrictive effect on cerebral vessels [10] but it is unclear whether A β levels are increased in the AD retina. Plaques and tangles have not been found in the human retina although A β has been isolated in aged human retinas [132]. Research on the retinas of AD transgenic mice has demonstrated A β plaques, hyperphosphorylated tau, increased microvascular deposition of A β and neuroinflammation [133-134]. A β immunotherapy in such transgenic mice has resulted in the clearance of retinal plaques but an increase in retinal amyloid angiopathy, identifying non-invasive retinal imaging as an alternative method for monitoring disease response to immunotherapy in these mice.

In addition to retinal venular constriction in human AD, Berisha *et al.* demonstrated significant thinning of the superior RNFL using optical coherence tomography (OCT, see Figs.

5 and 6). This region corresponds with the inferior visual field and these changes could explain the vision loss reported in this area in AD [48]. Other OCT studies have reported different patterns of RNFL loss (general, parapapillary and macular) [119, 135-136]. Iseri *et al.* found macular thinning in AD to be related to the severity of cognitive impairment [136]. Parisi *et al.* found RNFL thinning to be related to retinal dysfunction as revealed by abnormal pattern electroretinogram (PERG) responses [135]. Other studies have also found abnormal PERG responses in AD [137-138].

(Figure 5)

(Figure 6)

The loss of RNFL thickness in AD is linked to a depletion of retinal ganglion cells (RGC) and optic nerve axons as identified by histopathological studies [61, 139-143]. RGCs are the final common pathway that transfer visual information through the retinal nerve fibres and then the optic nerve into the brain (Fig. 7). A postmortem study by Blanks *et al.* [141] demonstrated a 25% decrease in RGC at the level of the foveal and parafoveal retina, while other studies have found no significant changes [144-145]. Further research is needed to confirm the RGC loss in AD, to establish any connection with A β in the retina and to confirm whether RGC loss is a cause of visual impairment in AD.

(Figure 7)

Retinal photography (Fig. 4) has also been used to identify RNFL abnormalities (nerve fiber loss) in AD [121-122], although one study indicated practical difficulties in using this approach for AD screening. The retinal nerve fibers are thick enough in the inner retina to make them visible in retinal photographs (Fig. 4), hence revealing areas of RNFL loss. While

Berisha *et al.* found reduced venular caliber in AD using a laser Doppler device, no retinal photography study has yet confirmed any retinal vascular changes in AD, possibly because such measurements using photography are complicated by issues of calibration and vessel boundary identification. Nevertheless, retinal photography does have the potential to detect RNFL and retinal vascular changes and hence needs to be pursued further in AD screening research.

AD is known to have a vascular component, with small-vessel disease, microinfarction and cerebral amyloid angiopathy (characterised by A β deposition in vessel walls) [146-147]. A β plaques as well as retinal microvascular deposition of A β have been identified in the retinas of AD transgenic mouse models [133] and could possibly be detected by non-invasive optical scattering [110], Raman spectroscopic or fluorescent tagging techniques. Given the homology between the retinal and cerebral microvasculatures [148], it is not unexpected that changes in the retinal vasculature might also occur in AD. Vascular topography, including the angles at which blood vessels bifurcate and the relationship between the widths of parent to daughter blood vessels at vascular junctions is optimized in healthy subjects in order to minimize shear stress across a vascular network [149-150]. Variations from the optimal geometrical topography are known to occur in particular vascular conditions [151-152]. Similar variations may occur in AD due to the disease's vascular component and hence are worthy of being explored with retinal photography.

Retinal photography and Scanning Laser Ophthalmoscopy (SLO) have both been used to demonstrate optic disc changes in AD, including optic disc pallor, pathologic disc cupping (hollowing-out), and thinning of the neuro-retinal rim [122-123]. Some of the ocular morphologies found in AD are also found in the eye disease glaucoma, specifically RNFL thinning, optic disc cupping and visual field loss. Glaucoma is second only to cataract as a

leading cause of blindness worldwide [153] and has ocular hypertension as its largest risk factor. The 5-fold higher chance of visual field defects and/or optic disc cupping found in AD has been interpreted as a higher occurrence rate of glaucoma in AD [124]. However, in this study, no AD participants had a family history of glaucoma, and ocular hypertension was not found in AD participants but was found in 7.5% of controls, reducing the likelihood that open-angle glaucoma was the cause. This is still in question though, with another study supporting the increased incidence of open-angle glaucoma in AD [154].

A greater than 10%/year decay in visual field and optic disc cupping were demonstrated in glaucoma patients who were later diagnosed with AD, whereas an average 3%/year decay in visual field was observed in glaucoma patients who did not develop AD, indicating that AD accelerates the progression of glaucoma symptoms [126]. However, increased rates of visual field defects and/or optic disc cupping have also been reported in Parkinson's disease [125] and since these changes are observed in the common eye disease glaucoma, they are unlikely to provide a test that has specificity for AD.

However, it is possible that investigations into these retinal changes in AD and glaucoma might yield interesting results about the pathogenesis of the diseases, as well as their treatment and monitoring. The similarities between the ocular effects of AD and glaucoma extend to changes observed in PERG recordings [127], the type of cells lost (large magnocellular RGC [61]) and possibly to the mechanism of RGC loss (apoptosis) [128-129]. In experimental glaucoma, A β co-localizes with RGC apoptosis and induces RGC apoptosis *in vivo* [130]. In addition, targeting the A β pathway with a β -secretase inhibitor, Congo red or A β -antibody has been found to be effective in treating experimental glaucoma (reducing RGC apoptosis) [130].

Chronic ocular hypertension (elevated intra-ocular pressure – IOP) has long been assumed to be a cause of RGC loss in glaucoma and has been shown to increase A β production in the rat retina [128]. Interestingly, in addition to its neuroprotective effects, a cholinesterase inhibitor used to treat AD has demonstrated dual therapeutic potential by reducing IOP in AD patients [155]. Current treatments for glaucoma are directed at reducing IOP, but evidence indicates that RGC loss still occurs in many glaucoma patients after successful IOP normalization, indicating that other mechanisms are involved.

In addition to glaucoma, A β has also been implicated in other retinal diseases such as age-related macular degeneration (AMD) [156-159]. This provides hope that these common neurodegenerative diseases (AD, Glaucoma and AMD) could be targeted simultaneously for treatment and monitoring, but reduces the likelihood of an A β -based, AD-specific biomarker in the retina. If A β changes in the retina are to be useful for AD screening, significant differences from the changes observed in retinal diseases must be identified. A β plaques as well as retinal microvascular deposition of A β have been identified in the retinas of AD transgenic mouse models [133]. If such changes occur in human AD, they could possibly be detected by non-invasive optical scattering [110], Raman spectroscopic or fluorescent tagging techniques. AD brain tissues exhibit visual and infrared excited auto-fluorescence (AF) [118]. Retinal AF changes are observed in a number of eye disorders including glaucoma, in which lipofuscin accumulation in parapapillary retinal pigment epithelial (RPE) cells causes AF changes [160]. Given the similarity of the retinal cell death process reported in glaucoma and AD [128-129], AF changes are also worthy of being investigated in the AD retina. It is hoped that these techniques could prove useful for non-invasive, early diagnosis of AD using the eye.

Conclusions

The terrible impact of AD, both on those directly affected and on society in general, creates a pressing need for better treatments. By the time a person is diagnosed with “probable AD” using current techniques, significant irreversible neuronal degeneration has already occurred. Therefore, research into better treatments must be paralleled by research into technologies to screen populations for AD, to identify cases before cognitive symptoms arise.

Ocular morphologies reported in AD give hope for a non-invasive, cost-effective screening test for AD. Evidence is accumulating in support of AD-related changes in the eye, but finding a sufficiently sensitive and specific ocular biomarker is proving to be a major challenge. Many reported ocular changes in AD also occur in other disorders. Optic disc changes, visual field defects and RNFL/retinal cell loss are also observed in the eye disease glaucoma. Retinal vessel widths are influenced by age and race and can be altered in many disorders. Similarly, pupil responses are influenced by age and eye color and are altered in many neurological disorders. Also, most studies into ocular morphology in AD have been limited by small participant numbers, few study groups and little relevant medical information on participants. Hence larger studies are required to confirm and investigate further these ocular changes in AD.

There is no doubt that the substantial diagnostic error rate in AD is confounding many aspects of AD research. A combination of new brain imaging techniques and CSF biomarkers has the potential to alleviate this problem, leading to more robust results in AD research, including ocular morphology. No studies have yet compared ocular morphology in AD to brain A β -loading or CSF protein changes, an approach that has the potential to shed light on

connections between ocular changes and AD. There are also many more ocular parameters left to investigate in AD.

Thus there remains scope for ocular changes to be utilised in AD screening or diagnostic purposes with greater sensitivity and specificity and at an earlier stage in the disease process. An ocular screening test for AD would benefit AD sufferers and researchers and possibly provide new insight into the molecular processes and genetic determinants of the disease. An ocular biomarker or biomarkers could turn out to be highly specific for AD, or to be a useful component in a multidisciplinary approach aimed at producing an earlier and more accurate diagnosis of Alzheimer's disease.

Bibliography

- [1] Brookmeyer R, Johnson E, Ziegler-Graham K, Arrighi HM (2007) Forecasting the global burden of Alzheimer's disease. *Alzheimer's and Dementia* **3**, 186-191.
- [2] Brookmeyer R, Gray S, Kawas C (1998) Projections of Alzheimer's disease in the United States and the public health impact of delaying disease onset. *Am J Public Health* **88**, 1337-1342.
- [3] Petersen RC, Stevens JC, Ganguli M, Tangalos EG, Cummings JL, DeKosky ST (2001) Practice parameter: Early detection of dementia: Mild cognitive impairment (an evidence-based review): Report of the Quality Standards Subcommittee of the American Academy of Neurology. *Neurology* **56**, 1133-1142.
- [4] Tiraboschi P, Hansen LA, Thal LJ, Corey-Bloom J (2004) The importance of neuritic plaques and tangles to the development and evolution of AD. *Neurology* **62**, 1984-1989.
- [5] Bates KA, Sohrabi HR, Rodrigues M, Beilby J, Dhaliwal SS, Taddei K, Criddle A, Wraith M, Howard M, Martins G, Paton A, Mehta P, Foster JK, Martins IJ, Lautenschlager NT, Mastaglia FL, Laws SM, Gandy SE, Martins RN (2009) Association of cardiovascular factors and Alzheimer's disease plasma amyloid-beta protein in subjective memory complainers. *J Alzheimers Dis* **17**, 305-318.
- [6] Shankar GM, Li S, Mehta TH, Garcia-Munoz A, Shepardson NE, Smith I, Brett FM, Farrell MA, Rowan MJ, Lemere CA, Regan CM, Walsh DM, Sabatini BL, Selkoe DJ

- (2008) Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat Med* **14**, 837-842.
- [7] Xu F, Davis J, Miao J, Previti ML, Romanov G, Ziegler K, Van Nostrand WE (2005) Protease nexin-2/amyloid beta-protein precursor limits cerebral thrombosis. *Proc Natl Acad Sci U S A* **102**, 18135-18140.
- [8] Turner PR, O'Connor K, Tate WP, Abraham WC (2003) Roles of amyloid precursor protein and its fragments in regulating neural activity, plasticity and memory. *Prog Neurobiol* **70**, 1-32.
- [9] Kang J, Lemaire HG, Unterbeck A, Salbaum JM, Masters CL, Grzeschik KH, Multhaup G, Beyreuther K, Muller-Hill B (1987) The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. *Nature* **325**, 733-736.
- [10] Suo Z, Humphrey J, Kundtz A, Sethi F, Placzek A, Crawford F, Mullan M (1998) Soluble Alzheimers beta-amyloid constricts the cerebral vasculature in vivo. *Neurosci Lett* **257**, 77-80.
- [11] Chan C-W, Dharmarajan A, Atwood CS, Huang X, Tanzi RE, Bush AI, Martins RN (1999) Anti-apoptotic action of Alzheimer ABeta. *ALZHEIMERS REPORTS* **2**, 113-119.
- [12] Hardy J, Cullen K (2006) Amyloid at the blood vessel wall. *Nat Med* **12**, 756-757.
- [13] Soccia SJ, Kirby JE, Washicosky KJ, Tucker SM, Ingelsson M, Hyman B, Burton MA, Goldstein LE, Duong S, Tanzi RE, Moir RD (2010) The Alzheimer's disease-associated amyloid beta-protein is an antimicrobial peptide. *PLoS One* **5**, e9505.
- [14] Butterfield DA, Lauderback CM (2002) Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and consequences involving amyloid beta-peptide-associated free radical oxidative stress. *Free Radic Biol Med* **32**, 1050-1060.
- [15] Lue LF, Kuo YM, Roher AE, Brachova L, Shen Y, Sue L, Beach T, Kurth JH, Rydel RE, Rogers J (1999) Soluble amyloid beta peptide concentration as a predictor of synaptic change in Alzheimer's disease. *Am J Pathol* **155**, 853-862.
- [16] Tanzi RE, Bertram L (2001) New frontiers in Alzheimer's disease genetics. *Neuron* **32**, 181-184.
- [17] Yin YI, Bassit B, Zhu L, Yang X, Wang C, Li YM (2007) {gamma}-Secretase Substrate Concentration Modulates the A β 42/A β 40 Ratio: IMPLICATIONS FOR ALZHEIMER DISEASE. *J Biol Chem* **282**, 23639-23644.
- [18] Ramsden M, Kotilinek L, Forster C, Paulson J, McGowan E, SantaCruz K, Guimaraes A, Yue M, Lewis J, Carlson G, Hutton M, Ashe KH (2005) Age-dependent neurofibrillary tangle formation, neuron loss, and memory impairment in a mouse model of human tauopathy (P301L). *J Neurosci* **25**, 10637-10647.
- [19] Santacruz K, Lewis J, Spire T, Paulson J, Kotilinek L, Ingelsson M, Guimaraes A, DeTure M, Ramsden M, McGowan E, Forster C, Yue M, Orne J, Janus C, Mariash A, Kuskowski M, Hyman B, Hutton M, Ashe KH (2005) Tau suppression in a neurodegenerative mouse model improves memory function. *Science* **309**, 476-481.
- [20] Spire TL, Orne JD, SantaCruz K, Pitstick R, Carlson GA, Ashe KH, Hyman BT (2006) Region-specific dissociation of neuronal loss and neurofibrillary pathology in a mouse model of tauopathy. *Am J Pathol* **168**, 1598-1607.
- [21] Bush AI (2002) Metal complexing agents as therapies for Alzheimer's disease. *Neurobiol Aging* **23**, 1031-1038.
- [22] Bush AI (2003) The metallobiology of Alzheimer's disease. *Trends Neurosci* **26**, 207-214.

- [23] Schenk D, Barbour R, Dunn W, Gordon G, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Liao Z, Lieberburg I, Motter R, Mutter L, Soriano F, Shopp G, Vasquez N, Vandever C, Walker S, Wogulis M, Yednock T, Games D, Seubert P (1999) Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse. *Nature* **400**, 173-177.
- [24] Parker MH, Chen R, Conway KA, Lee DH, Luo C, Boyd RE, Nortey SO, Ross TM, Scott MK, Reitz AB (2002) Synthesis of (-)-5,8-dihydroxy-3R-methyl-2R-(dipropylamino)-1,2,3,4-tetrahydronaphthalene: an inhibitor of beta-amyloid(1-42) aggregation. *Bioorg Med Chem* **10**, 3565-3569.
- [25] Doody RS, Gavrilova SI, Sano M, Thomas RG, Aisen PS, Bachurin SO, Seely L, Hung D (2008) Effect of dimebon on cognition, activities of daily living, behaviour, and global function in patients with mild-to-moderate Alzheimer's disease: a randomised, double-blind, placebo-controlled study. *Lancet* **372**, 207-215.
- [26] Medivation, Press release 3 March 2010. Pfizer and Medivation announce results from two Phase 3 studies in Dimebon (latrepirdine*) Medivation website [online], <http://investors.medivation.com/releasedetail.cfm?ReleaseID=448818>, Accessed June 2010.
- [27] Lazarov O, Robinson J, Tang YP, Hairston IS, Korade-Mirnic Z, Lee VM, Hersh LB, Sapolsky RM, Mirnic K, Sisodia SS (2005) Environmental enrichment reduces Abeta levels and amyloid deposition in transgenic mice. *Cell* **120**, 701-713.
- [28] Wilson RS, Mendes De Leon CF, Barnes LL, Schneider JA, Bienias JL, Evans DA, Bennett DA (2002) Participation in cognitively stimulating activities and risk of incident Alzheimer disease. *JAMA* **287**, 742-748.
- [29] Cirrito JR, Yamada KA, Finn MB, Sloviter RS, Bales KR, May PC, Schoepp DD, Paul SM, Mennerick S, Holtzman DM (2005) Synaptic activity regulates interstitial fluid amyloid-beta levels in vivo. *Neuron* **48**, 913-922.
- [30] Buckner RL, Snyder AZ, Shannon BJ, LaRossa G, Sachs R, Fotenos AF, Sheline YI, Klunk WE, Mathis CA, Morris JC, Mintun MA (2005) Molecular, structural, and functional characterization of Alzheimer's disease: evidence for a relationship between default activity, amyloid, and memory. *J Neurosci* **25**, 7709-7717.
- [31] Schaffer C, Donlon P (1983) Medical causes of psychiatric symptoms in the elderly. *Clinical Gerontologist* **1**, 3-18.
- [32] Thal LJ, Kantarci K, Reiman EM, Klunk WE, Weiner MW, Zetterberg H, Galasko D, Pratico D, Griffin S, Schenk D, Siemers E (2006) The role of biomarkers in clinical trials for Alzheimer disease. *Alzheimer Dis Assoc Disord* **20**, 6-15.
- [33] Sunderland T, Linker G, Mirza N, Putnam KT, Friedman DL, Kimmel LH, Bergeson J, Manetti GJ, Zimmermann M, Tang B, Bartko JJ, Cohen RM (2003) Decreased beta-amyloid1-42 and increased tau levels in cerebrospinal fluid of patients with Alzheimer disease. *JAMA* **289**, 2094-2103.
- [34] Fagan AM, Mintun MA, Mach RH, Lee SY, Dence CS, Shah AR, LaRossa GN, Spinner ML, Klunk WE, Mathis CA, DeKosky ST, Morris JC, Holtzman DM (2006) Inverse relation between in vivo amyloid imaging load and cerebrospinal fluid Abeta42 in humans. *Ann Neurol* **59**, 512-519.
- [35] Rowe CC, Ng S, Ackermann U, Gong SJ, Pike K, Savage G, Cowie TF, Dickinson KL, Maruff P, Darby D, Smith C, Woodward M, Merory J, Tochon-Danguy H, O'Keefe G, Klunk WE, Mathis CA, Price JC, Masters CL, Villemagne VL (2007) Imaging beta-amyloid burden in aging and dementia. *Neurology* **68**, 1718-1725.

- [36] Morris JC, Roe CM, Grant EA, Head D, Storandt M, Goate AM, Fagan AM, Holtzman DM, Mintun MA (2009) Pittsburgh Compound B Imaging and Prediction of Progression From Cognitive Normality to Symptomatic Alzheimer Disease. *Arch Neurol* **66**, 1469-1475.
- [37] Selkoe DJ (2001) Alzheimer's disease: genes, proteins, and therapy. *Physiol Rev* **81**, 741-766.
- [38] Bu G (2009) Apolipoprotein E and its receptors in Alzheimer's disease: pathways, pathogenesis and therapy. *Nat Rev Neurosci* **10**, 333-344.
- [39] Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* **261**, 921-923.
- [40] Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, Pahwa JS, Moskva V, Dowzell K, Williams A, Jones N, Thomas C, Stretton A, Morgan AR, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Morgan K, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Love S, Kehoe PG, Hardy J, Mead S, Fox N, Rossor M, Collinge J, Maier W, Jessen F, Schurmann B, van den Bussche H, Heuser I, Kornhuber J, Wiltfang J, Dichgans M, Frolich L, Hampel H, Hull M, Rujescu D, Goate AM, Kauwe JS, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Muhleisen TW, Nothen MM, Moebus S, Jockel KH, Klopp N, Wichmann HE, Carrasquillo MM, Pankratz VS, Younkin SG, Holmans PA, O'Donovan M, Owen MJ, Williams J (2009) Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet* **41**, 1088-1093.
- [41] Lambert JC, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, Combarros O, Zelenika D, Bullido MJ, Tavernier B, Letenneur L, Bettens K, Berr C, Pasquier F, Fievet N, Barberger-Gateau P, Engelborghs S, De Deyn P, Mateo I, Franck A, Helisalmi S, Porcellini E, Hanon O, de Pancorbo MM, Lendon C, Dufouil C, Jaillard C, Leveillard T, Alvarez V, Bosco P, Mancuso M, Panza F, Nacmias B, Bossu P, Piccardi P, Annoni G, Seripa D, Galimberti D, Hannequin D, Licastro F, Soininen H, Ritchie K, Blanche H, Dartigues JF, Tzourio C, Gut I, Van Broeckhoven C, Alperovitch A, Lathrop M, Amouyel P (2009) Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet* **41**, 1094-1099.
- [42] Bertram L, Tanzi RE (2008) Thirty years of Alzheimer's disease genetics: the implications of systematic meta-analyses. *Nat Rev Neurosci* **9**, 768-778.
- [43] Villemagne VL, Pike KE, Darby D, Maruff P, Savage G, Ng S, Ackermann U, Cowie TF, Currie J, Chan SG, Jones G, Tochon-Danguy H, O'Keefe G, Masters CL, Rowe CC (2008) Abeta deposits in older non-demented individuals with cognitive decline are indicative of preclinical Alzheimer's disease. *Neuropsychologia* **46**, 1688-1697.
- [44] Pike KE, Savage G, Villemagne VL, Ng S, Moss SA, Maruff P, Mathis CA, Klunk WE, Masters CL, Rowe CC (2007) Beta-amyloid imaging and memory in non-demented individuals: evidence for preclinical Alzheimer's disease. *Brain* **130**, 2837-2844.
- [45] Simonsen AH, McGuire J, Podust VN, Davies H, Minthon L, Skoog I, Andreasen N, Wallin A, Waldemar G, Blennow K (2008) Identification of a novel panel of cerebrospinal fluid biomarkers for Alzheimer's disease. *Neurobiol Aging* **29**, 961-968.

- [46] Katz B, Rimmer S (1989) Ophthalmologic manifestations of Alzheimer's disease. *Surv Ophthalmol* **34**, 31-43.
- [47] Sadun AA, Borchert M, DeVita E, Hinton DR, Bassi CJ (1987) Assessment of visual impairment in patients with Alzheimer's disease. *Am J Ophthalmol* **104**, 113-120.
- [48] Trick GL, Trick LR, Morris P, Wolf M (1995) Visual field loss in senile dementia of the Alzheimer's type. *Neurology* **45**, 68-74.
- [49] Whittaker KW, Burdon MA, Shah P (2002) Visual field loss and Alzheimer's disease. *Eye (Lond)* **16**, 206-208.
- [50] Pache M, Smeets CH, Gasio PF, Savaskan E, Flammer J, Wirz-Justice A, Kaiser HJ (2003) Colour vision deficiencies in Alzheimer's disease. *Age Ageing* **32**, 422-426.
- [51] Cogan DG (1987) Alzheimer syndromes. *Am J Ophthalmol* **104**, 183-184.
- [52] Cronin-Golomb A, Sugiura R, Corkin S, Growdon JH (1993) Incomplete achromatopsia in Alzheimer's disease. *Neurobiology of Aging* **14**, 471-477.
- [53] Lakshminarayanan V, Lagrave J, Kean ML, Dick M, Shankle R (1996) Vision in dementia: contrast effects. *Neurol Res* **18**, 9-15.
- [54] Crow RW, Levin LB, LaBree L, Rubin R, Feldon SE (2003) Sweep visual evoked potential evaluation of contrast sensitivity in Alzheimer's dementia. *Invest Ophthalmol Vis Sci* **44**, 875-878.
- [55] Nissen MJ, Corkin S, Buonanno FS, Growdon JH, Wray SH, Bauer J (1985) Spatial vision in Alzheimer's disease. General findings and a case report. *Arch Neurol* **42**, 667-671.
- [56] Mendola JD, Cronin-Golomb A, Corkin S, Growdon JH (1995) Prevalence of visual deficits in Alzheimer's disease. *Optom Vis Sci* **72**, 155-167.
- [57] Schlotterer G, Moscovitch M, Crapper-McLachlan D (1984) Visual processing deficits as assessed by spatial frequency contrast sensitivity and backward masking in normal ageing and Alzheimer's disease. *Brain* **107 (Pt 1)**, 309-325.
- [58] Mielke R, Kessler J, Fink G, Herholz K, Heiss WD (1995) Dysfunction of visual cortex contributes to disturbed processing of visual information in Alzheimer's disease. *Int J Neurosci* **82**, 1-9.
- [59] Morrison JH, Hof PR, Bouras C (1991) An anatomic substrate for visual disconnection in Alzheimer's disease. *Ann N Y Acad Sci* **640**, 36-43.
- [60] Gilmore GC, Wenk HE, Naylor LA, Koss E (1994) Motion Perception and Alzheimer's Disease. *Journal of Gerontology* **49**, P52-P57.
- [61] Sadun AA, Bassi CJ (1990) Optic nerve damage in Alzheimer's disease. *Ophthalmology* **97**, 9-17.
- [62] Fletcher WA, Sharpe JA (1988) Smooth pursuit dysfunction in Alzheimer's disease. *Neurology* **38**, 272-277.
- [63] Mendez MF, Tomsak RL, Remler B (1990) Disorders of the visual system in Alzheimer's disease. *J Clin Neuroophthalmol* **10**, 62-69.
- [64] Leuba G (1995) Pathology of subcortical visual centres in relation to cortical degeneration in Alzheimer's disease. *Neuropathology and applied neurobiology* **21**, 410.
- [65] McKee AC, Au R, Cabral HJ, Kowall NW, Seshadri S, Kubilus CA, Drake J, Wolf PA (2006) Visual association pathology in preclinical Alzheimer disease. *J Neuropathol Exp Neurol* **65**, 621-630.
- [66] Bentley P, Driver J, Dolan RJ (2008) Cholinesterase inhibition modulates visual and attentional brain responses in Alzheimer's disease and health. *Brain* **131**, 409-424.

- [67] Herholz K, Weisenbach S, Zundorf G, Lenz O, Schroder H, Bauer B, Kalbe E, Heiss WD (2004) In vivo study of acetylcholine esterase in basal forebrain, amygdala, and cortex in mild to moderate Alzheimer disease. *Neuroimage* **21**, 136-143.
- [68] Nobili L, Sannita WG (1997) Cholinergic modulation, visual function and Alzheimer's dementia. *Vision Res* **37**, 3559-3571.
- [69] Scinto LF, Daffner KR, Dressler D, Ransil BI, Rentz D, Weintraub S, Mesulam M, Potter H (1994) A potential noninvasive neurobiological test for Alzheimer's disease. *Science* **266**, 1051-1054.
- [70] Idiaquez J, Alvarez G, Villagra R, San Martin RA (1994) Cholinergic supersensitivity of the iris in Alzheimer's disease. *J Neurol Neurosurg Psychiatry* **57**, 1544-1545.
- [71] Pomara N, Sitaram N (1995) Detecting Alzheimer's disease. *Science* **267**, 1579-1580; author reply 1580-1571.
- [72] Iijima A, Haida M, Ishikawa N, Ueno A, Minamitani H, Shinohara Y (2003) Re-evaluation of tropicamide in the pupillary response test for Alzheimer's disease. *Neurobiol Aging* **24**, 789-796.
- [73] Gomez-Tortosa E, del Barrio A, Jimenez-Alfaro I (1996) Pupil response to tropicamide in Alzheimer's disease and other neurodegenerative disorders. *Acta Neurol Scand* **94**, 104-109.
- [74] Grunberger J, Linzmayer L, Walter H, Rainer M, Masching A, Pezawas L, Saletu-Zyhlarz G, Stohr H, Grunberger M (1999) Receptor test (pupillary dilatation after application of 0.01% tropicamide solution) and determination of central nervous activation (Fourier analysis of pupillary oscillations) in patients with Alzheimer's disease. *Neuropsychobiology* **40**, 40-46.
- [75] Kalman J, Kanka A, Magloczky E, Szoke A, Jardenhazy T, Janka Z (1997) Increased mydriatic response to tropicamide is a sign of cholinergic hypersensitivity but not specific to late-onset sporadic type of Alzheimer's dementia. *Biol Psychiatry* **41**, 909-911.
- [76] Robles A, Tourino R, Sesar A, Suarez P, Noya M (1996) [Experience with pupil tropicamide test in Alzheimer's disease]. *Rev Neurol* **24**, 65-68.
- [77] Kono K, Miyao M, Ishihara S, Takagi A, Ikari H, Suzuki Y, Iguchi A (1996) [Hypersensitivity in the pupil dilation response to a cholinergic antagonist in patients with Alzheimer's disease and Down's syndrome]. *Nippon Ronen Igakkai Zasshi* **33**, 829-834.
- [78] Arai H, Terajima M, Nakagawa T, Higuchi S, Mochizuki H, Sasaki H (1996) Pupil dilatation assay by tropicamide is modulated by apolipoprotein E epsilon 4 allele dosage in Alzheimer's disease. *Neuroreport* **7**, 918-920.
- [79] Tohgi H, Abe T, Hashiguchi K, Saheki M, Takahashi S (1994) Remarkable reduction in acetylcholine concentration in the cerebrospinal fluid from patients with Alzheimer type dementia. *Neurosci Lett* **177**, 139-142.
- [80] Hou RH, Samuels ER, Raisi M, Langley RW, Szabadi E, Bradshaw CM (2006) Why patients with Alzheimer's disease may show increased sensitivity to tropicamide eye drops: role of locus coeruleus. *Psychopharmacology (Berl)* **184**, 95-106.
- [81] FitzSimon JS, Waring SC, Kokmen E, McLaren JW, Brubaker RF (1997) Response of the pupil to tropicamide is not a reliable test for Alzheimer disease. *Arch Neurol* **54**, 155-159.
- [82] Kardon RH (1998) Drop the Alzheimer's drop test. *Neurology* **50**, 588-591.

- [83] Caputo L, Casartelli M, Perrone C, Santori M, Annoni G, Vergani C (1998) The 'eye test' in recognition of late-onset Alzheimer's disease. *Arch Gerontol Geriatr* **27**, 171-177.
- [84] Fridh M, Havelius U, Elofsson G, Hindfelt B (1996) The pupillary response to tropicamide in Alzheimer's disease. *Acta Ophthalmol Scand* **74**, 276-279.
- [85] Growdon JH, Graefe K, Tennis M, Hayden D, Schoenfeld D, Wray SH (1997) Pupil dilation to tropicamide is not specific for Alzheimer disease. *Arch Neurol* **54**, 841-844.
- [86] Kurz A, Marquard R, Fremke S, Leipert KP (1997) Pupil dilation response to tropicamide: a biological test for Alzheimer's disease? *Pharmacopsychiatry* **30**, 12-15.
- [87] Marx JL, Kumar SR, Thach AB, Kiat-Winarko T, Frambach DA (1995) Detecting Alzheimer's disease. *Science* **267**, 1577; author reply 1580-1571.
- [88] Reitner A, Baumgartner I, Thuile C, Baradaran Dilmaghani R, Ergun E, Kaminski S, Lukas J, Dal Bianco P (1997) The mydriatic effect of tropicamide and its diagnostic use in Alzheimer's disease. *Vision Res* **37**, 165-168.
- [89] Loupe DN, Newman NJ, Green RC, Lynn MJ, KK WI, Geis TC, Edelhauser HF (1996) Pupillary response to tropicamide in patients with Alzheimer disease. *Ophthalmology* **103**, 495-503.
- [90] Treloar AJ, Assin M, Macdonald AJ (1996) Pupillary response to topical tropicamide as a marker for Alzheimer's disease. *Br J Clin Pharmacol* **41**, 256-257.
- [91] Granholm E, Morris S, Galasko D, Shults C, Rogers E, Vukov B (2003) Tropicamide effects on pupil size and pupillary light reflexes in Alzheimer's and Parkinson's disease. *Int J Psychophysiol* **47**, 95-115.
- [92] Higuchi S, Matsushita S, Hasegawa Y, Muramatsu T, Arai H, Hayashida M (1997) Apolipoprotein E epsilon 4 allele and pupillary response to tropicamide. *Am J Psychiatry* **154**, 694-696.
- [93] Sacks B, Smith S (1989) People with Down's syndrome can be distinguished on the basis of cholinergic dysfunction. *J Neurol Neurosurg Psychiatry* **52**, 1294-1295.
- [94] Higuchi S, Matsushita S, Hasegawa Y, Muramatsu T, Arai H (1997) Pupillary response to tropicamide in Japanese patients with alcoholic dementia, Alzheimer's disease, and vascular dementia. *Exp Neurol* **144**, 199-201.
- [95] Den Heijer JC, Bollen WL, Reulen JP, van Dijk JG, Kramer CG, Roos RA, Buruma OJ (1988) Autonomic nervous function in Huntington's disease. *Arch Neurol* **45**, 309-312.
- [96] Meyer S, Gibb T, Jurkovich GJ (1993) Evaluation and significance of the pupillary light reflex in trauma patients. *Ann Emerg Med* **22**, 1052-1057.
- [97] Steinhauer SR, Hakerem G (1992) The pupillary response in cognitive psychophysiology and schizophrenia. *Ann N Y Acad Sci* **658**, 182-204.
- [98] Van Diemen HA, Van Dongen MM, Nauta JJ, Lanting P, Polman CH (1992) Pupillary light reflex latency in patients with multiple sclerosis. *Electroencephalogr Clin Neurophysiol* **82**, 213-219.
- [99] Loewenfeld IE (1999) *The pupil: Anatomy, physiology, and clinical applications*.
- [100] Fotiou DF, Brozou CG, Haidich AB, Tsiptsios D, Nakou M, Kabitsi A, Giantselidis C, Fotiou F (2007) Pupil reaction to light in Alzheimer's disease: evaluation of pupil size changes and mobility. *Aging Clin Exp Res* **19**, 364-371.

- [101] Fotiou F, Fountoulakis KN, Tsolaki M, Goulas A, Palikaras A (2000) Changes in pupil reaction to light in Alzheimer's disease patients: a preliminary report. *Int J Psychophysiol* **37**, 111-120.
- [102] Prettyman R, Bitsios P, Szabadi E (1997) Altered pupillary size and darkness and light reflexes in Alzheimer's disease. *J Neurol Neurosurg Psychiatry* **62**, 665-668.
- [103] Fotiou DF, Brozou CG, Tsiptsios DJ, Fotiou A, Kabitsi A, Nakou M, Giantselidis C, Goula A (2007) Effect of age on pupillary light reflex: evaluation of pupil mobility for clinical practice and research. *Electromyogr Clin Neurophysiol* **47**, 11-22.
- [104] Hanson SR, Hasan A, Smith DL, Smith JB (2000) The major in vivo modifications of the human water-insoluble lens crystallins are disulfide bonds, deamidation, methionine oxidation and backbone cleavage. *Exp Eye Res* **71**, 195-207.
- [105] Spector A (1995) Oxidative stress-induced cataract: mechanism of action. *FASEB J* **9**, 1173-1182.
- [106] Goldstein LE, Muffat JA, Cherny RA, Moir RD, Ericsson MH, Huang X, Mavros C, Coccia JA, Faget KY, Fitch KA, Masters CL, Tanzi RE, Chylack LT, Jr., Bush AI (2003) Cytosolic beta-amyloid deposition and supranuclear cataracts in lenses from people with Alzheimer's disease. *Lancet* **361**, 1258-1265.
- [107] Yoneda S, Hara H, Hirata A, Fukushima M, Inomata Y, Tanihara H (2005) Vitreous fluid levels of beta-amyloid((1-42)) and tau in patients with retinal diseases. *Jpn J Ophthalmol* **49**, 106-108.
- [108] Moncaster JA, Pineda R, Moir RD, Lu S, Burton MA, Ghosh JG, Ericsson M, Soscia SJ, Mocofanescu A, Folkert RD, Robb RM, Kuszak JR, Clark JI, Tanzi RE, Hunter DG, Goldstein LE (2010) Alzheimer's disease amyloid-beta links lens and brain pathology in Down syndrome. *PLoS One* **5**, e10659.
- [109] Frederikse PH, Garland D, Zigler JS, Jr., Piatigorsky J (1996) Oxidative stress increases production of beta-amyloid precursor protein and beta-amyloid (A β) in mammalian lenses, and A β has toxic effects on lens epithelial cells. *J Biol Chem* **271**, 10169-10174.
- [110] Goldstein LE, Moir R, Lu S, Fu L, Chadwick O, Arnett E, Ericsson M, Klunk W, Mathis C, Chylack LT, Jr., Clark J, Tanzi R, Moncaster JA (2006) Non-invasive early detection of beta-amyloid molecular pathology by quasi-elastic light scattering in vivo. *Alzheimer's & dementia* **2**, S133.
- [111] Ansari RR, Datiles MB, 3rd (1999) Use of dynamic light scattering and Scheimpflug imaging for the early detection of cataracts. *Diabetes Technol Ther* **1**, 159-168.
- [112] Datiles MB, 3rd, Ansari RR, Reed GF (2002) A clinical study of the human lens with a dynamic light scattering device. *Exp Eye Res* **74**, 93-102.
- [113] Datiles MB, 3rd, Ansari RR, Suh KI, Vitale S, Reed GF, Zigler JS, Jr., Ferris FL, 3rd (2008) Clinical detection of precatactous lens protein changes using dynamic light scattering. *Arch Ophthalmol* **126**, 1687-1693.
- [114] Sudworth CD (2006) Advances in Raman spectroscopy for the diagnosis of Alzheimer's disease. *Proc SPIE* **6093**, 139-146.
- [115] Hanlon EB, Perelman LT, Vitkin EI, Greco FA, McKee AC, Kowall NW (2008) Scattering differentiates Alzheimer disease in vitro. *Opt Lett* **33**, 624-626.
- [116] Hanlon EB, Itzkan I, Dasari RR, Feld MS, Ferrante RJ, McKee AC, Lathi D, Kowall NW (1999) Near-infrared fluorescence spectroscopy detects Alzheimer's disease in vitro. *Photochem Photobiol* **70**, 236-242.

- [117] Archer JKJ, Sudworth CD, Williams R, How T, Stone N, Mann D, Black RA (2007) Improvements in Alzheimer' Disease Diagnosis using Principal Component Analysis (PCA) in Combination with Raman Spectroscopy. *Proc SPIE* **6628**, 37.
- [118] Zipfel WR, Williams RM, Christie R, Nikitin AY, Hyman BT, Webb WW (2003) Live tissue intrinsic emission microscopy using multiphoton-excited native fluorescence and second harmonic generation. *Proc Natl Acad Sci U S A* **100**, 7075-7080.
- [119] Paquet C, Boissonnot M, Roger F, Dighiero P, Gil R, Hugon J (2007) Abnormal retinal thickness in patients with mild cognitive impairment and Alzheimer's disease. *Neurosci Lett* **420**, 97-99.
- [120] Berisha F, Feke GT, Trempe CL, McMeel JW, Schepens CL (2007) Retinal abnormalities in early Alzheimer's disease. *Invest Ophthalmol Vis Sci* **48**, 2285-2289.
- [121] Hedges TR, 3rd, Perez Galves R, Speigelman D, Barbas NR, Peli E, Yardley CJ (1996) Retinal nerve fiber layer abnormalities in Alzheimer's disease. *Acta Ophthalmol Scand* **74**, 271-275.
- [122] Tsai CS (1991) Optic nerve head and nerve fiber layer in Alzheimer's disease. *Archives of ophthalmology* **109**, 199.
- [123] Danesh-Meyer HV, Birch H, Ku JY, Carroll S, Gamble G (2006) Reduction of optic nerve fibers in patients with Alzheimer disease identified by laser imaging. *Neurology* **67**, 1852-1854.
- [124] Bayer AU, Ferrari F, Erb C (2002) High Occurrence Rate of Glaucoma among Patients with Alzheimer's Disease. *European Neurology* **47**, 165-168.
- [125] Bayer AU, Keller ON, Ferrari F, Maag KP (2002) Association of glaucoma with neurodegenerative diseases with apoptotic cell death: Alzheimer's disease and Parkinson's disease. *Am J Ophthalmol* **133**, 135-137.
- [126] Bayer AU, Ferrari F (2002) Severe progression of glaucomatous optic neuropathy in patients with Alzheimer's disease. *Eye* **16**, 209-212.
- [127] Neshet R, Trick GL (1991) The pattern electroretinogram in retinal and optic nerve disease. A quantitative comparison of the pattern of visual dysfunction. *Doc Ophthalmol* **77**, 225-235.
- [128] McKinnon SJ, Lehman DM, Kerrigan-Baumrind LA, Merges CA, Pease ME, Kerrigan DF, Ransom NL, Tahzib NG, Reitsamer HA, Levkovitch-Verbin H, Quigley HA, Zack DJ (2002) Caspase activation and amyloid precursor protein cleavage in rat ocular hypertension. *Invest Ophthalmol Vis Sci* **43**, 1077-1087.
- [129] Yin H, Chen L, Chen X, Liu X (2008) Soluble amyloid beta oligomers may contribute to apoptosis of retinal ganglion cells in glaucoma. *Med Hypotheses* **71**, 77-80.
- [130] Guo L, Salt TE, Luong V, Wood N, Cheung W, Maass A, Ferrari G, Russo-Marie F, Sillito AM, Cheetham ME, Moss SE, Fitzke FW, Cordeiro MF (2007) Targeting amyloid-beta in glaucoma treatment. *Proc Natl Acad Sci U S A* **104**, 13444-13449.
- [131] Sun C, Sun (2009) Retinal Vascular Caliber: Systemic, Environmental, and Genetic Associations. *Survey of Ophthalmology* **54**, 74.
- [132] Loffler KU (1995) Immunoreactivity against tau, amyloid precursor protein, and beta-amyloid in the human retina. *Investigative ophthalmology & visual science* **36**, 24.
- [133] Liu B, Rasool S, Yang Z, Glabe CG, Schreiber SS, Ge J, Tan Z (2009) Amyloid-peptide vaccinations reduce {beta}-amyloid plaques but exacerbate vascular deposition and inflammation in the retina of Alzheimer's transgenic mice. *Am J Pathol* **175**, 2099-2110.

- [134] Perez SE, Lumayag S, Kovacs B, Mufson EJ, Xu S (2009) Beta-amyloid deposition and functional impairment in the retina of the APP^{swe}/PS1^{DeltaE9} transgenic mouse model of Alzheimer's disease. *Invest Ophthalmol Vis Sci* **50**, 793-800.
- [135] Parisi V, Restuccia R, Fattapposta F, Mina C, Bucci MG, Pierelli F (2001) Morphological and functional retinal impairment in Alzheimer's disease patients. *Clin Neurophysiol* **112**, 1860-1867.
- [136] Iseri PK, Altinas O, Tokay T, Yuksel N (2006) Relationship between cognitive impairment and retinal morphological and visual functional abnormalities in Alzheimer disease. *J Neuroophthalmol* **26**, 18-24.
- [137] Katz B, Rimmer S, Iragui V, Katzman R (1989) Abnormal pattern electroretinogram in Alzheimer's disease: evidence for retinal ganglion cell degeneration? *Ann Neurol* **26**, 221-225.
- [138] Trick GL, Barris MC, Bickler-Bluth M (1989) Abnormal pattern electroretinograms in patients with senile dementia of the Alzheimer type. *Ann Neurol* **26**, 226-231.
- [139] Blanks JC, Hinton DR, Sadun AA, Miller CA (1989) Retinal ganglion cell degeneration in Alzheimer's disease. *Brain Res* **501**, 364-372.
- [140] Blanks JC, Schmidt SY, Torigoe Y, Porrello KV, Hinton DR, Blanks RH (1996) Retinal pathology in Alzheimer's disease. II. Regional neuron loss and glial changes in GCL. *Neurobiol Aging* **17**, 385-395.
- [141] Blanks JC, Torigoe Y, Hinton DR, Blanks RH (1996) Retinal pathology in Alzheimer's disease. I. Ganglion cell loss in foveal/parafoveal retina. *Neurobiol Aging* **17**, 377-384.
- [142] Hinton DR, Sadun AA, Blanks JC, Miller CA (1986) Optic-nerve degeneration in Alzheimer's disease. *N Engl J Med* **315**, 485-487.
- [143] Sadun AA, Bassi CJ (1990) The visual system in Alzheimer's disease. *Res Publ Assoc Res Nerv Ment Dis* **67**, 331-347.
- [144] Curcio CA, Drucker DN (1993) Retinal ganglion cells in Alzheimer's disease and aging. *Ann Neurol* **33**, 248-257.
- [145] Davies DC, McCoubrie P, McDonald B, Jobst KA (1995) Myelinated axon number in the optic nerve is unaffected by Alzheimer's disease. *Br J Ophthalmol* **79**, 596-600.
- [146] Ravona-Springer R, Davidson M, Noy S (2003) The role of cardiovascular risk factors in Alzheimer's disease. *CNS Spectr* **8**, 824-833.
- [147] Thal DR, Ghebremedhin E, Orantes M, Wiestler OD (2003) Vascular pathology in Alzheimer disease: correlation of cerebral amyloid angiopathy and arteriosclerosis/lipohyalinosis with cognitive decline. *J Neuropathol Exp Neurol* **62**, 1287-1301.
- [148] Patton N, Aslam T, Macgillivray T, Pattie A, Deary IJ, Dhillon B (2005) Retinal vascular image analysis as a potential screening tool for cerebrovascular disease: a rationale based on homology between cerebral and retinal microvasculatures. *J Anat* **206**, 319-348.
- [149] Zamir M, Brown N (1982) Arterial branching in various parts of the cardiovascular system. *Am J Anat* **163**, 295-307.
- [150] Zamir M, Medeiros JA (1982) Arterial branching in man and monkey. *J Gen Physiol* **79**, 353-360.
- [151] Chapman N, Dell'omo G, Sartini MS, Witt N, Hughes A, Thom S, Pedrinelli R (2002) Peripheral vascular disease is associated with abnormal arteriolar diameter relationships at bifurcations in the human retina. *Clin Sci (Lond)* **103**, 111-116.

- [152] Stanton AV, Wasan B, Cerutti A, Ford S, Marsh R, Sever PP, Thom SA, Hughes AD (1995) Vascular network changes in the retina with age and hypertension. *J Hypertens* **13**, 1724-1728.
- [153] Resnikoff S, Pascolini D, Etya'ale D, Kocur I, Pararajasegaram R, Pokharel GP, Mariotti SP (2004) Global data on visual impairment in the year 2002. *Bull World Health Organ* **82**, 844-851.
- [154] Tamura H, Kawakami H, Kanamoto T, Kato T, Yokoyama T, Sasaki K, Izumi Y, Matsumoto M, Mishima HK (2006) High frequency of open-angle glaucoma in Japanese patients with Alzheimer's disease. *Journal of the Neurological Sciences* **246**, 79-83.
- [155] Estermann S, Daepf GC, Cattapan-Ludewig K, Berkhoff M, Frueh BE, Goldblum D (2006) Effect of oral donepezil on intraocular pressure in normotensive Alzheimer patients. *J Ocul Pharmacol Ther* **22**, 62-67.
- [156] Johnson LV, Leitner WP, Rivest AJ, Staples MK, Radeke MJ, Anderson DH (2002) The Alzheimer's A beta -peptide is deposited at sites of complement activation in pathologic deposits associated with aging and age-related macular degeneration. *Proc Natl Acad Sci U S A* **99**, 11830-11835.
- [157] Luibl V, Isas JM, Kaye R, Glabe CG, Langen R, Chen J (2006) Drusen deposits associated with aging and age-related macular degeneration contain nonfibrillar amyloid oligomers. *The Journal of Clinical Investigation* **116**, 378-385.
- [158] Anderson DH, Talaga KC, Rivest AJ, Barron E, Hageman GS, Johnson LV (2004) Characterization of [beta] amyloid assemblies in drusen: the deposits associated with aging and age-related macular degeneration. *Experimental Eye Research* **78**, 243-256.
- [159] Dentchev T, Milam AH, Lee VMY, Trojanowski JQ, Dunaief JL (2003) Amyloid- β is found in drusen from some age-related macular degeneration retinas, but not in drusen from normal retinas¹ Edited by Hans E. Grossniklaus, MD. *American journal of ophthalmology* **136**, 787.
- [160] Viestenz A, Langenbacher A, Mardin CY (2006) [Parapapillary autofluorescence as indicator for glaucoma]. *Klin Monatsbl Augenheilkd* **223**, 315-320.

Table 1: Reported Ocular Changes in AD

Part of the eye	Reported Ocular Changes in AD	Journal (Year)	Ref.	N(AD,Control)
Pupil	Enhanced pupil response to cholinergic drops	Science (1994)	[69]	19,32
		Neuroreport (1996)	[78]	25,24
		J Neurol Neurosurg Psych (1994)	[70]	26,23
		Neurobiol Aging (2003)	[72]	14,30
		Acta Neurol Scand (1996)	[73]	24,50
		Neuropsychobiology (1999)	[74]	29,29
		Biol Psychiatry (1997)	[75]	67,80
		Rev Neurol (1996)	[76]	10,20
		Nippon Ronen Igakkai Zasshi (1996)	[77]	53,29
Pupil	Altered pupil flash response	Aging Clin Exp Res (2007)	[100]	23,23
		Int J Psychophysiol (2000)	[101]	10,5
		Int J Psychophysiol (2003)	[91]	15,30
		J Neurol Neurosurg Psych (1997)	[102]	9,9
Lens	Aggregation of A β , Supra-nuclear cataract	Lancet (2003)	[106]	9,8
Retina	Narrow retinal veins and decreased venular blood flow	Invest Ophthalmol Vis Sci (2007)	[120]	9,8
Retina	Retinal Nerve Fiber Layer (RNFL) thinning	Invest Ophthalmol Vis Sci (2007)	[120]	9,8
		Neurosci Lett (2007)	[119]	26,38
Retina	RNFL abnormalities and cell loss	Acta Neurol Scand (1996)	[121]	26,23
		Archives of Ophthalmology (1991)	[122]	26,30
		Neurology (2006)	[123]	40,50
Retina	Abnormal pattern electroretinogram (PERG)	Ann Neurol (1989)	[137]	6,6
		Ann Neurol (1989)	[138]	13,30
Optic Disc	Optic disc pallor, pathologic disc cupping, and thinning of the neuro-retinal rim	Acta Neurol Scand (1996)	[121]	26,23
		Archives of Ophthalmology (1991)	[122]	30,32
		Neurology (2006)	[123]	40,50

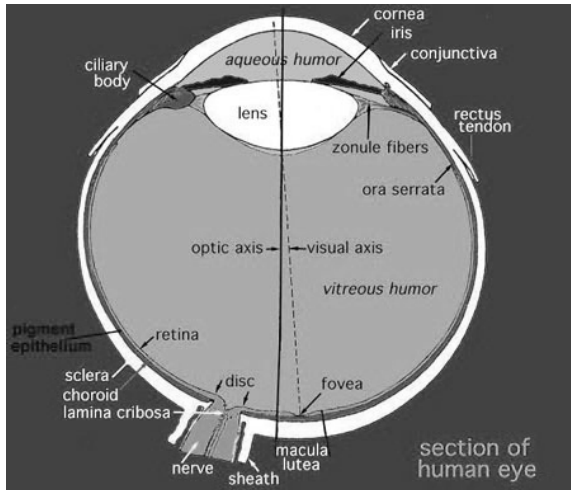
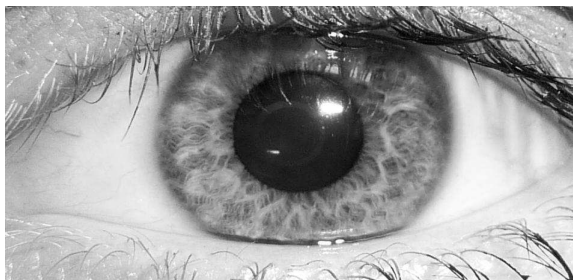
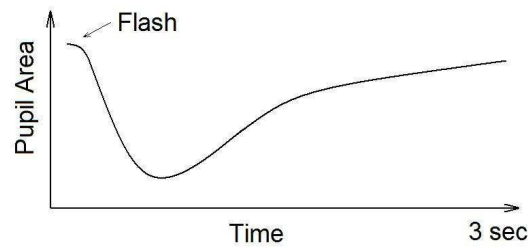


Figure 1: Cross-section of the human eye



(a)



(b)

Figure 2. (a) The human eye. The pupil is the central transparent aperture (appearing as black), surrounded by the iris. (b) The graph of a pupil flash response, showing the pupil contraction resulting from a bright flash of white light.

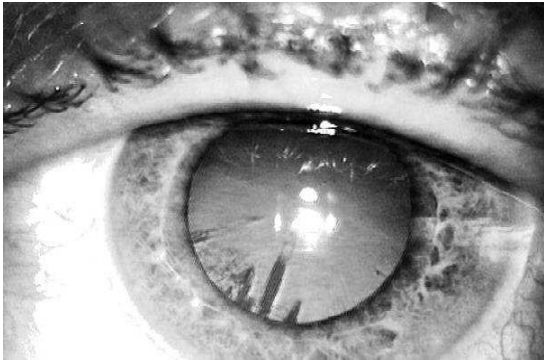


Figure 3. Retro-illumination photograph of a lens with cataract, pupil dilated with tropicamide eye drops. A different type of cataract has been linked with Alzheimer's disease.

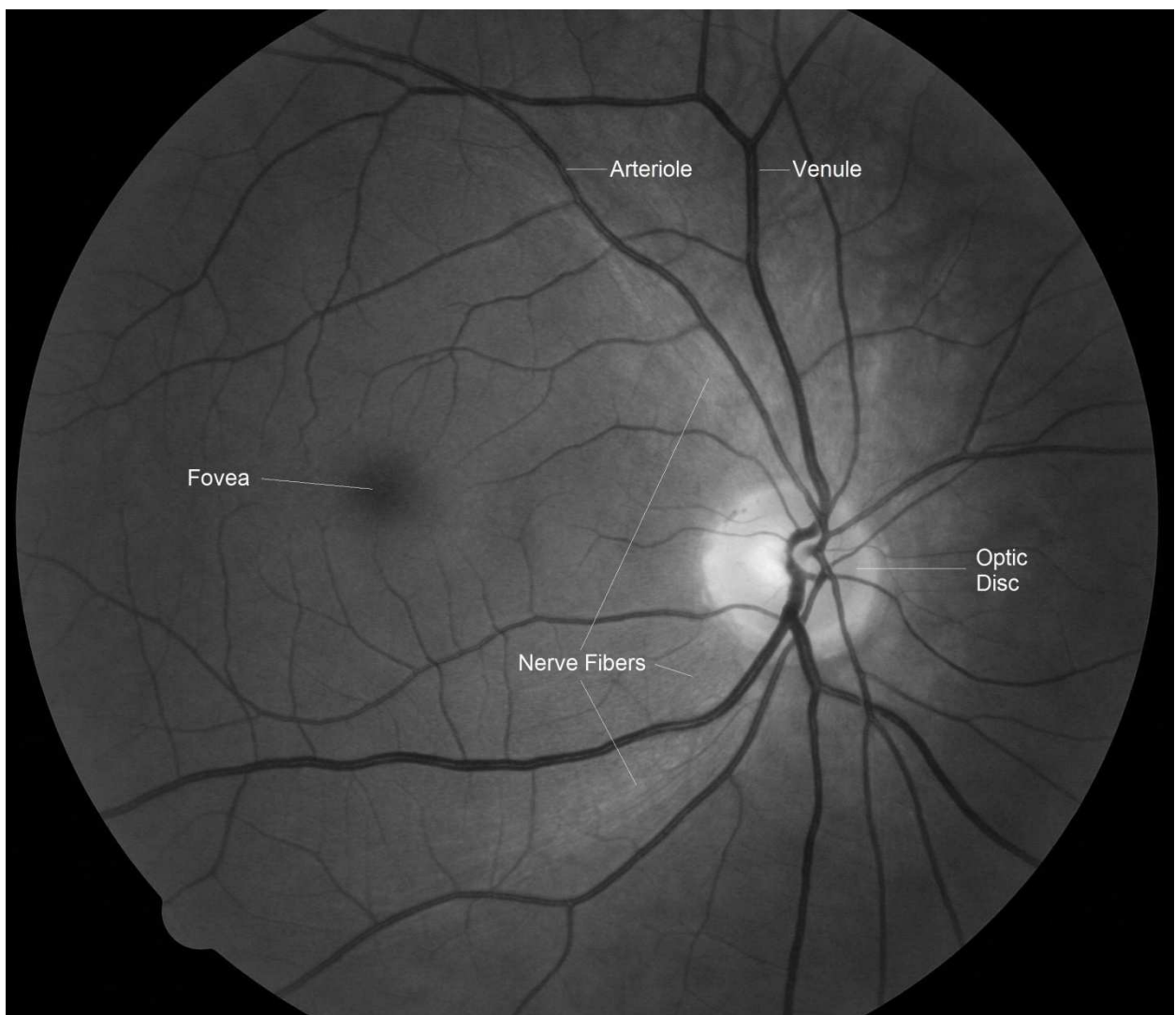


Figure 4. Digital retinal photograph displaying the optic disc in centre, with retinal arterioles and venules (darker) and lightly opaque retinal nerve fibers coursing to the optic disc.

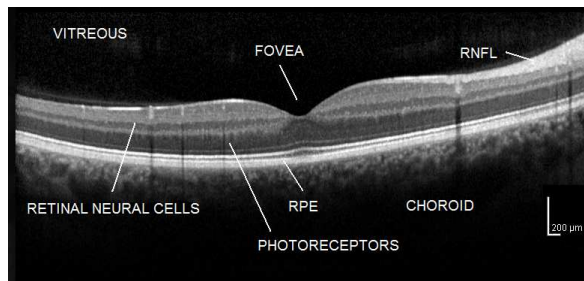


Figure 5. OCT scan showing the retinal layers around the fovea. The layer closest to the vitreous humour is the retinal nerve fiber layer (RNFL) which contains fibers emerging from the retinal ganglion cells below. Also just beneath the RNFL is the retinal vasculature (evident from the vertical shadows cast in this OCT scan). Beneath the retinal ganglion cells are the bipolar, amacrine and horizontal cells, followed by a layer of photoreceptor cells. The photoreceptor cells are nourished by the deeper retinal pigment epithelium and a rich posterior vascular layer called the choroid. *OCT scan courtesy of Chris Barry, Lions Eye Institute, Perth, Australia.*

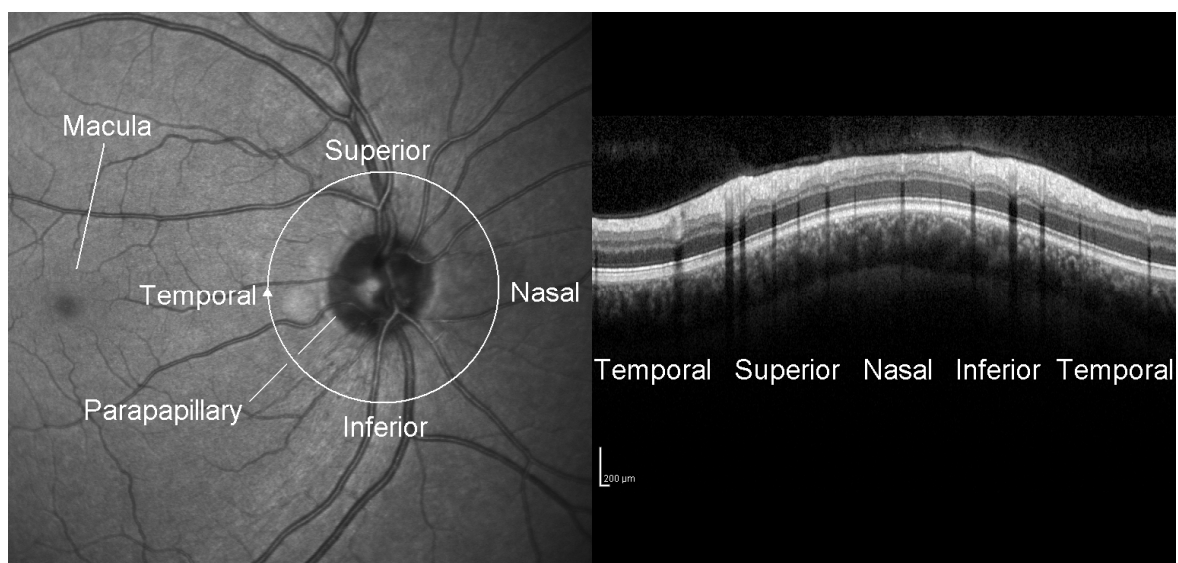


Figure 6. OCT scan circling the optic disc. The image on the right shows the retinal layers detected in an OCT scan traversing a circular path around the optic disc, as illustrated in the retinal photograph on the left. The RNFL is thickest in the superior and inferior quadrants. RNFL studies in AD have had varying results, indicating superior, general, macular or parapapillary thinning. *OCT scan courtesy of Chris Barry, Lions Eye Institute, Perth, Australia.*

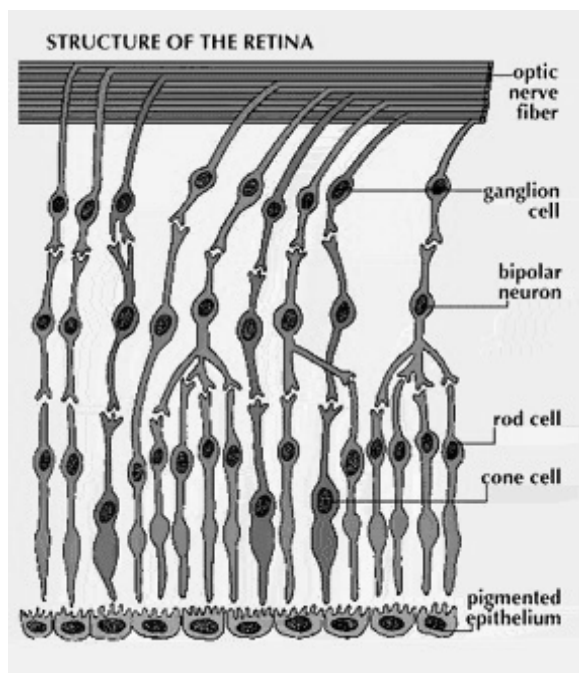


Figure 7: Layers of cells in the retina. Light must pass through the retinal nerve fiber layer and retinal neural cells (ganglion, bipolar, etc.) before reaching the photoreceptor cells (rods and cones).